

## ADVANCE TRAINING IN BIOTECHNOLOGY

The overall goal of the training program in biotechnology can lead to a multitude of careers in botany, genetics, medicine and biotechnology. While entry-level positions can be achieved with a bachelor's degree, greater levels of education afford more opportunities - specifically with regards to research and teaching opportunities.



### Scope of training :

- Research & Development
- Molecular Diagnostic Lab
- Food & Beverage Industry
- Pharmaceutical Industry
- Herbal & Nutraceuticals Industry
- Forensic Labs

### Advantage -

- Industry Oriented Design
- Spirit of the training is to make you employable
- Rigorous Hands on Learning
- Capacity Building
- State of art research facility
- Flexibility of Time & Working Hours

**Duration :** 500 Hours ( 4 - 6 Months ) **Timings :** Monday - Saturday ( 9 A.M to 5 P.M )

### Important Note:

- **This training program have the potential to make you employable**
- **As a part of our training program, we may suggest your name to our clients against available vacancy**
- **This training is limited to skill development only and we do not assure you for employment or campus interview**
- **Our prime motive is “Bridging Gap Between Industry & Academia”**

# TRAINING SYLLABUS

## LAB SAFETY, PROCEDURES & REGULATIONS

Lab safety and Procedures , Record Maintenance, Handling of Equipments , Sterilisation Techniques , Preparation of Chemical & Reagents Discussion of ethical, legal, and social issues involved in molecular biotechnology. IPR & Biotechnology, Regulatory, Ethics & Guidelines For Genetic Research

## MOLECULAR GENETICS

### UNIT II :- Extraction , Quantification and Purification of DNA , RNA , mRNA, cDNA

Extraction of Nucleic Acid - Both DNA & RNA Quantitative & Qualitative Analysis of Nucleic Acid - DNA & RNA Electrophoresis , Gel Docking or imaging . Quantitative analysis by spectrometer – For DNA – Measure Absorbance at 260 & 280 nm For RNA – Quantitative Analysis by Orisinol Method , mRNA Purification & cDNA

**Preparation Note** - You will be suggested to extract best quality nucleic acid for further use in PCR , Real Time PCR , cDNA Preparation , Sequencing , Microarray . All nucleic acid should pass strict quality check .

### UNIT III :- Bio Informatics Tools & Techniques :

Primer Designing, Vectors , Selection of Restriction Sites, Virtual PCR, Bioinformatics tools & Techniques , Gel Analysis Software , Vector Selection Software Real Time PCR - Primer designing Tool , Software to run Real Time PCR , Data Analysis Microarray - Microarray Instrument Control and Data Analysis Software

### UNIT IV :-Handling of PCR , Real Time PCR & Microarray Platform

**PCR** - Preparation of reaction mixture and its safety for cross contamination , Optimisation of PCR Reaction – Melting Point , GC Content , Concentrations and Cycles , Sample run , Data Analysis.

**Real Time PCR** - Reaction setup for real time PCR, selection of quantification – relative or absolute , Melting curve analysis , SYBR Green Assay , Sample run.

**Microarray Platform** - Microarray Instrument Control , cDNA Preparation , Hybridisation with used chip or array for learning and data analysis .

### UNIT V - Expression Studies & Applications

Genetic Expression Studies through PCR – 16S rDNA Analysis, Conventional PCR Method , Nested PCR , PCR Multiplexing etc. mRNA Purification , cDNA Preparation , Real Time PCR assay and its different applications in molecular genetics.

### UNIT VI :- Genetic Toxicology & Applications

**Bioassay Development** : Basics of Chromatography : Column Chromatography , Thin Layer Chromatography , HPLC , Gas Chromatography & Mass Spectrometry

**Bio separation Assays by HPLC** : Sample preparation , Introduction to separation techniques , SPME Separations , Method Development for mutagenesis assay, Analysis of genetic mutagenesis assay by HPLC

**DNA Methylation Studies** - Bisulfites modification of DNA , Bisulfites modification in nanogram quantities of DNA , DNA Methylation specific PCR assay .

## **UNIT VII - Re-Combinant DNA Technology**

Isolation of pUC18 plasmid from TOP10-pUC18 E coli cells , Restriction digestion of pUC 18 and  $\lambda$  DNA , Purifying pUC18/Hind III/ EcoR I digest by gel elution , Ligating the linearized plasmid - pUC18 and the insert  $\lambda$ DNA. , Preparation of competent cells , Transformation of TOP10 cells with the pUC18- $\lambda$ DNA ligated product Colony PCR : To amplify the inserted  $\lambda$ DNA digest in pUC18 vector

## **UNIT VIII – cDNA Library Construction**

Extraction of RNA , Purification of mRNA through Oligo-dT Column Chromatography, cDNA Construction , Incorporation of cDNA into a vector , Cloning of cDNAs

## **PROTEOMICS STUDY**

### **UNIT I- Protein Extraction & Protein Assay**

Protein Extraction, Acid Base Equilibrium, pH, Buffer System, Charge, pI and pKa Value, Quantitative determination of biomolecule, mini scale bacterial protein extraction, protein extraction from plant source or other biological source

### **UNIT II - Protein Estimation & Quantitation**

Protein estimation by Lowry's Assay / Biuret Assay / Bradford's Method / BCA Method , Densitometry Analysis of Protein

### **UNIT III - Protein Purification by FPLC, LPLC & HPLC**

Purification of Protein by Affinity Chromatography ( IMAC / GST / Sepharose etc) , Ion Exchange Chromatography , Size Exclusion and Hydrophobic Purification, Desalting, Dialysis, Ultrafiltration and centrifugation.

### **UNIT IV - Protein Characterisation**

Protein Characterisation by SDS-PAGE, Native Page, Zymography, Iso Electric Focussing, 2-D Electrophoresis, Western Blot, Staining by Coomassie Blue, Supra Ruby, Deep Purple Protein Fingerprinting, Immunodiffusion Assay

### **UNIT V - Amino Acid Analysis by Chromatography**

Sample preparation, Hydrolysis, Derivatization, Separation of derivatized amino acids, Data Interpretation and calculation

### **UNIT VI - Protein Bioinformatics**

Analysis of 2-D Data, Peptide Mass Fingerprinting Data Analysis ( LC-MS MALDI-TOF ), Homology Modelling, Molecular Docking

## **MICROBIOLOGY & METAGENOMICS**

### **UNIT I - Isolation & Enumeration of Microorganism**

Microbial Growth- Isolation & Plating Techniques, Single colony isolation, Determination of microbial count, Growth Curve Analysis

## **UNIT II - Bio-Chemical Characterisation For Preliminary Screening**

Basic biochemical testing; IMVIC, Reducing Sugar, Gram Staining, Morphology, Triple Sugar Iron Agar , Starch Hydrolysis, Lipid Hydrolysis, Casein Hydrolysis and many more , **In-silico studies of the positive or negative data for microbial identification**

## **UNIT III - Meta-Genomics & NGS Data Analysis**

**Next Generation Sequence Data Analysis ( amplicon based )**, Amplicon Target Population Structure, Meta-genome Shotgun Processing, Q.C of meta-genomics Sequence Data, **Real Time PCR Primer Design**, Q-PCR Data Handling,

## **UNIT IV - 16S / 23S / 28S rRNA Analysis**

DNA Extraction & Quantitation for Metagenomic Analysis, PCR and its Optimisation, Thermostable DNA Polymerases; PCR Melting Curve Analysis , Amplification of Genomic DNA, PCR Multiplexing , Elution of PCR amplicon , Purification of PCR Product for Sequencing

## **UNIT V - Functional Meta-genomics**

Quantitative or Real Time PCR can be coupled with 16S rRNA species specific primers to assess species population

## **ANALYTICAL & CHROMATOGRAPHY TECHNIQUES**

### **UNIT I - Extraction & Sample Preparations**

**Extraction Procedures** - Different extraction methods for volatile , semi volatile and non volatile samples through digestion , soxhlet extraction , distillation , Vacuum Rotary Evaporator , Solid Phase Micro Extraction etc.

**Sample Preparation For Chromatography** - Solid Phase Extraction, Solid Phase Micro Extraction, Ultra Sonication etc.

### **UNIT II : QUALITY CONTROL AND QUALITY ASSURANCE**

**Quality Control Procedures** - Physical and Chemical Analysis , Initial Method Validation , On Going Method Validation , Laboratory Blanks , Duplicate Determinations , Calibrations , Q.C. Calculations , Q.C. Charts etc. Operation and Calibration of Meters : pHMeter (Hanna , Thermo ) , Conductivity Meter , Dissolved Oxygen Meter , Spectrophotometer , Pipettes , Turbidity Meter etc.

### **UNIT III - LEARNING ON HPLC , GC , SPECTROSCOPY AND OTHER TECHNIQUES**

**Analysis by HPLC** – Basics of HPLC - Sample Preparation , Gradient Making , Parts of HPLC , Troubleshooting and Maintenance , Operating Procedure of HPLC , Run the sample in HPLC . Data Analysis

**Analysis by Gas Chromatography** – Basics of GC - Sample Preparation , Parts of GC , Troubleshooting and Maintenance , Operating Procedure of GC , Run the sample in GC , Data Analysis

#### **Analysis by Spectroscopy & TLC –**

Basics of Spectroscopy - Sample Preparation , Calibration and Calibration Curve , Recovery Percentage etc. Analysis by Thin Layer Chromatography – Sample Preparation , Solvent Selection , Spray Selection , Visualisation and analysis of TLC bands.

## **UNIT IV – Analysis of Samples**

- Analysis of adulterants in Food Samples
- Analysis of adulterants in Beverages & Liquor
- Analysis of Sample of Forensic Importance
- Analysis of Pharmaceutical Drug

## **WHO MAY JOIN ?**

**Indian** Aspirants From Biotechnology , Microbiology , Biochemistry , Life Science , Chemistry , Pharmacy ,Forensic Science , Food Science etc.

**Fee Structure** : Rs 40,000 /- ( Payable in Three Instalments )

**Duration** : 500 Hours ( 4 - 6 Months ) **Timings** : Monday - Saturday ( 9 A.M to 5 P.M )

## HOW TO APPLY –


### Details of Documents For Registration :

1. Any identity proof along with University / College Identity Card / Aadhar Card etc.
2. Filled **Registration form** with photograph ( Given in Last Page of Brochure )
2. **Registration fee** will be Rs 1000 / - paid through cheque or on line payment

### How to pay Registration Fee Off Line ( Those Who Send Documents by Post ) :

1. Cheque or D.D will be in favour of “ **Allele Life Sciences Private Limited**”

### On Line Payment :

Payment By Internet Banking	Scan UPI Code
<b>Beneficiary Name - Allele Life Sciences Private Limited</b> <b>Account Number - 61071508494</b> <b>IFSC Code - SBIN0031811</b> <b>Bank Name - State Bank of India</b> <b>Bank Address - SBI, 14/15, Sector-18, Noida, UP - 201301</b>  <b>Or Pay Through UPI / BHIM App</b> <b>UPI Address - allelelifesciences@upi</b>	

### How to send document :

Those who pay through cheque send all documents at following address :

#### **Allele Life Sciences Pvt. Ltd.**

C - 59 , Sector - 10 , Noida  
Uttar Pradesh - 201301 , IN  
M : + 91-9891179928

Those who opt on Line registration send scan copy of all documents and receipt of online payment at : [allelelifesciences@gmail.com](mailto:allelelifesciences@gmail.com)

**Note :** We will send confirmation within specified time through e.mail or remind us.

## Registration Form

Name of Training Program :

Expected Date of Joining :

Candidate Details :

Name: .....

Father's Name: .....

Address : .....

Contact No : .....

Email: .....

Institution : .....

Qualification : .....

### Terms & Conditions :

1. The admission to training / internship programs will be confirmed after the payment of registration fee along with documents.
2. The registration fee deposited is completely non refundable.
3. The industrial training fee includes the cost of chemical , reagents and study material costs.
4. I will deposit the service charges as decided by the company at the time of joining date of training program.
5. Students have to bear their own boarding/lodging /conveyance charges. We facilitate students in finding proper paying guest arrangements.
6. The trainees will have to bring their own lab coat and wear them all the time in the campus.
7. Trainees are selected on first come first serve basis
8. Trainees will maintain adequate discipline inside the lab premises.
9. Company will not be responsible for any medical, legal issues during the internship tenure.

### DECLARATION

I \_\_\_\_\_ from \_\_\_\_\_  
hereby declare that all statement/information given in the application form are true to the best of my knowledge and belief . I will strictly abide by the norms/lab etiquette during the training

Signature

Place: \_\_\_\_\_

Date: \_\_\_\_\_

**For office use only**

## Instruments Capabilities

**Our State of art facility** is located in Industrial Area of Noida (NCR) . The lab / research facility is Total : 6000 Sq Feet

<b>Affymatrix &amp; Agilent Microarray Platform</b>	Gene Expression Studies, Biomarker, Sequencing
<b>Real Time PCR ( ABI )</b>	Gene Expression, Sequence Detection
<b>PCR ( ABI, Biorad , Eurofins ) - 5 in numbers</b>	Amplification of nucleic acids
<b>Bioanalyser &amp; Spectrophotometer</b>	Quantification of Nucleic Acids
<b>Gel Documentation System</b>	Visualisation of Nucleic Acids, PCR Products etc.
<b>Electrophoresis &amp; Power Supply ( Biorad ) - 7 Sets</b>	Separation of Nucleic Acids & Other Arrays
<b>DNA Concentrator ( Thermo Speedvac )</b>	Nucleic Acid Extraction
<b>Centrifuge, High Speed Centrifuge - 8 Nos</b>	Sample Preparation
<b>PCR Station and other accessories</b>	

<b>Biorad Profinia Affinity Chromatography</b>	Affinity Chromatography - IMAC, GST, Antibody
<b>Biorad Biologic Low Pressure Chromatography</b>	Size Exclusion, Ion Exchange, Affinity etc.
<b>Preparative HPLC ( Thermo ) , Agilent 1100</b>	Bulk Protein Purification & Analysis
<b>GE Amersham 2-D Electrophoresis System</b>	Protein Characterisation
<b>Immunoblot, SDS-PAGE , Biorad HV Powerpac</b>	Visualisation of Nucleic Acids, PCR Products etc.
<b>Mass Spectrometry , ELISA, Immunoassay</b>	Protein Identification
<b>Cryo Preservation Facility &amp; Common Facility</b>	Sample Storage & Preparation

<b>Agilent HPLC System - PDA, FLD &amp; ECD Detector</b>	Separation and analysis of molecules
<b>Agilent GC with FID &amp; FPD Detectors</b>	Separation and analysis of molecules
<b>Thermo Prep HPLC with Dual Pump &amp; UV-Vis</b>	Bulk Purification & Analysis
<b>Shimadzu GC with FID &amp; NPD Detector</b>	Separation and analysis of molecules
<b>Triple Quad GC-MS System ( Agilent )</b>	Analysis of Semi Volatile & Volatile Compound
<b>LC-MS-MS ( API Sciex )</b>	Analysis of Non Volatile Compound
<b>Varian Carry Spectrophotometer</b>	Analytical Tool for various purpose
<b>Thermo Helios Spectrophotometer</b>	Analytical Tool for various purpose
<b>Vacuum Rotary Evaporator ( Buchi )</b>	Sample Preparation



## Other Analytical Chemistry Equipments :

Refractometer , Flame Photometer ( Toshniwal), Karl Fisher Titrator (Sistrionics), Potentiometer, Polarimeter , Tintometer ,Viscometer , Kjeldahl Distillation Unit , Kjeldahl Digestion Unit , Ion Selective for Fluoride Analysis ( Thermo Orion ) , Nephelometer , Soxhlet Extraction , Rotatory Vacuum Evaporator with chiller , etc.



**Microbiology & Cell Culture Facility :** Vertical Laminar Air Flow ( 4x2x2 ) , Horizontal Laminar Air Flow ( 2x2x2 ) B.O.D. Incubator ( Julabo ) , CO<sub>2</sub> Incubator ( Jauan ) , Orbital Incubator Shaker, UV Chamber , Incubator, Colony Counter , Colorimeter , Muffle Furnace , Hot Air Oven , Desiccators, Binocular Microscopes and , Lypholizer

**Biochemistry / Organic Synthesis Chemistry Lab :** Spectrophotometer ( Thermo Heleus Alpha ) , Analytical Balance ( Sartorius ) , Ph Meter ( Thermo Orion ) , Ion Selective (Thermo Orion) , Conductivity Meter ( Thermo Orion ) , Dissolved Oxygen Meter ( Thermo Orion ) , Turbidity Meter, Autoclaves, Hot Air Oven , Hot Plate , Magnetic Stirrers , Pipette Washer , Shaking Machine , Water Bath , Colorimeter , Flame Photometer , etc.

**Lab Water Purification :** Millipore Milli Q System

**Clinical Biology Lab :** Haematology Analyser , Automatic Immunoassay, Haematology HPLC Biorad Variant II



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